

Enhanced Hypoxia-Inducible Factor (HIF)-1 α Stability Induced by 5-Hydroxymethyl-2-Furfural (5-HMF) Contributes to Protection against Hypoxia

Yun-Ling He,^{1*} Ming-Ming Li,^{1,2*} Li-Ying Wu,¹ Tong Zhao,¹ Yao Di,¹ Xin Huang,¹ Xue-Feng Ding,¹ Kui-Wu Wu,¹ Ming Fan,^{1,2} and Ling-Ling Zhu¹

¹Department of Cognitive Science, Beijing Institute of Basic Medical Sciences, Beijing, China; and ²Beijing Institute for Brain Disorders, Beijing, China

We first reported the role of 5-hydroxymethyl-2-furfural (5-HMF) against hypoxia. Here, we studied the mechanism by using oxygen-dependent degradation domain (ODD)-Luc mice, which are a useful model to probe the stabilization of hypoxia-inducible factor 1 α (HIF-1 α). Compared with three other compounds that have been reported to have a role in stabilizing HIF-1 α , 5-HMF caused stronger bioluminescence, which is indicative of HIF-1 α stability in the brain and kidney of ODD-Luc mice. We further demonstrated that the HIF-1 α protein accumulated in response to 5-HMF in the brains and kidneys of these mice, as well as in PC12 cells. Additionally, 5-HMF promoted the nuclear translocation of HIF-1 α and the transcriptional activity of HIF-1, which was evaluated by detecting vascular endothelial growth factor (VEGF) mRNA expression. These results suggest that 5-HMF stabilized HIF-1 α and increased its activity. Considering the role of proline hydroxylases (PHDs) in negatively regulating HIF-1 α stability, we explored whether 5-HMF interacts with the substrates and cofactors of PHDs, such as 2-oxoglutarate (2-OG), Fe²⁺ and vitamin C (VC), which affects the activity of PHDs. The result revealed that 5-HMF did not interact with Fe²⁺ or 2-OG but interacted with VC. This interaction was confirmed by subsequent experiments, in which 5-HMF entered into cells and reduced the VC content. The enhanced stability of HIF-1 α by 5-HMF was reversed by VC supplementation, and the improved survival of mice caused by 5-HMF under hypoxia was abrogated by VC supplementation. Thus, we demonstrated for the first time that 5-HMF increases HIF-1 α stability by reducing the VC content, which mediates the protection against hypoxia.

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INTRODUCTION

Hypoxia is particularly damaging in the brain, where an oxygen deficiency can lead to stroke, cerebral palsy and epilepsy (1–4). For the threat of hypoxia,

prevention is far more important than treatment. In our recent study, we identified 5-hydroxymethyl-2-furfural (5-HMF) as a potential preventive drug against hypoxic diseases (5,6).

5-HMF is a product of the Maillard reaction and exists in plants, foods and beverages that contain carbohydrates, such as the roots of *Polygonum multiflorum*, dried fruits, bread, fruit juices, milk and coffee (7,8). The beneficial roles of 5-HMF include antioxidant activity, the inhibition of red blood cell sickling and the amelioration of hemorheology (4,9–12). Among these benefits, the role of antisickling, as indicated by the high affinity of 5-HMF for sickle cell hemoglobin, has been granted a U.S. patent (patent number 7119208). We first reported the role of 5-HMF in protecting against hypoxia in 2011 (5,6). Nevertheless, the underlying mechanisms have not yet been elucidated. In this study, we demonstrated for the first time that 5-HMF stabilizes hypoxia-inducible factor 1 α (HIF-1 α) through an interaction with VC, which mediates the protection of 5-HMF against hypoxia.

*Y-LH and M-ML contributed equally to this work.

Address correspondence to Ling-Ling Zhu, Department of Cognitive Science, Beijing Institute of Basic Medical Sciences, 27 Taiping Road, Beijing, 100850 P.R. China. Phone: +8610-66931315; Fax: +8610-68213039; E-mail: linglingzhu@hotmail.com; or Li-Ying Wu, Department of Cognitive Science, Beijing Institute of Basic Medical Sciences, 27 Taiping Road, Beijing, 100850 P.R. China. Phone: +8610-66931316; Fax: +8610-68213039; E-mail: liyiwu_china@163.com; or Ming Fan, Department of Cognitive Science, Beijing Institute of Basic Medical Sciences, 27 Taiping Road, Beijing, 100850 P.R. China. Phone: +8610-66932333; Fax: +8610-68213039; E-mail: fanmingchina@126.com.

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HIF-1 α is a subunit of hypoxia-inducible factor 1 (HIF-1) and determines HIF-1 activity. Under normoxia, HIF-1 α is extremely unstable and is quickly degraded by the ubiquitin-proteasome system. Proline hydroxylases (PHDs) (also known as HIF prolyl hydroxylases [HPH]) use O₂ and 2-oxoglutarate (2-OG) (also called α -ketoglutarate) as substrates and Fe²⁺ and vitamin C (VC) (also called ascorbic acid) as cofactors hydroxylate prolyl sites at residues 402 and 564 in the oxygen-dependent degradation domain (ODD) of HIF-1 α . Then, the tumor suppressor protein von Hippel-Lindau (VHL), which is an E3 ubiquitin ligase complex, recognizes hydroxylated HIF-1 α , and after polyubiquitination by ubiquitins, HIF-1 α is marked for destruction and is ultimately degraded by hydroxylases in proteasomes (13–15). Thus, it can be inferred that the reduction of O₂, 2-OG, Fe²⁺ or VC will inhibit the activities of PHDs and then elevate HIF-1 α stability. For example, under hypoxia, HIF-1 α becomes stable because of a lack of O₂ as a substrate of PHDs and, therefore, accumulates its protein. Subsequently, HIF-1 α translocates from the cytoplasm into the nucleus and heterodimerizes with constitutively expressed HIF-1 β , which is another HIF-1 subunit, to form the heterodimeric transcription factor HIF-1. HIF-1 binds to a conserved hypoxia-response element (HRE), which contains the core nucleotide sequence 5'-(A/G) CGTG-3' in the promoter region of its target genes, to activate the transcription of HIF-1 target genes. HIF-1 target genes include vascular endothelial growth factor (*VEGF*), erythropoietin (*EPO*), glucose transporters (*GLUTs*), *BCL2*/adenovirus E1B 19 kd-interacting protein 3 (*BNIP3*)/*BNIP3*-like (*BNIP3L*), miRNAs, and so on, which mediate the adaptation to hypoxic stress (16–18).

From the above description, we can easily draw an inference that the stability of HIF-1 α determines the hypoxic adaptation. Therefore, drugs that target HIF-1 α have the potential to prevent hypoxic diseases. Unfortunately, there is a lack of

drugs to stabilize HIF-1 α under normoxia and to prevent hypoxic injury. In our previous studies, we confirmed the protective role of 5-HMF against hypoxic injury in mice and in ECV304 cells (5,6). However, whether the role is correlated to HIF-1 α stability is unknown. In this study, using ODD-Luc transgenic mice to evaluate agents that stabilize HIF-1 α , we demonstrated for the first time that 5-HMF stabilizes HIF-1 α under normoxia by reducing the VC content, and this effect is relieved by VC supplementation. Thus, 5-HMF has the potential to be developed as a drug to prevent hypoxic diseases in the future.

MATERIALS AND METHODS

Materials

5-HMF, ferulic acid, coumarin, quercetin, CoCl₂ and ascorbate oxidase were purchased from Sigma-Aldrich (St. Louis, MO, USA). Luciferin was obtained from Promega (Madison, WI, USA). The HIF-1 α mouse monoclonal antibody was purchased from Novus Biologicals (Littleton, CO, USA). The mouse β -actin monoclonal antibody was acquired from Sigma-Aldrich. The rabbit Pro564-OH polyclonal antibody was purchased from Cell Signaling Technology (Boston, MA, USA). The Alexa 594 goat anti-mouse secondary antibody and 4',6-diamidino-2-phenylindole (DAPI) were Molecular Probes products (Thermo Fisher Scientific, Waltham, MA, USA).

Animals

The ODD-Luc mice were acquired from The Jackson Laboratory (Bar Harbor, ME, USA). This mouse model expresses the oxygen-dependent degradation (ODD) domain of HIF-1 α fused to the firefly luciferase gene. Fusing the ODD domain to luciferase renders the luciferase protein sensitive to oxygen-dependent hydroxylation and to subsequent ubiquitin-dependent degradation under normal physiological circumstances. The advantage of ODD-Luc mice is that these mice can provide a rapid

noninvasive measurement of possible agonists of HIF-1 activity (19).

C57BL/6J male mice were purchased from the Animal Central of Academy of Military Medical Science (Beijing, China) and were used for survival experiments under hypoxia conditions. The animal protocol was approved by the Institutional Animal Care and Use Committee of the Academy of Military Medical Science and was consistent with the National Institutes of Health (NIH) *Guide for the Care and Use of Laboratory Animals* (NIH publications no. 85-23) (20).

Cell Culture

The PC12 cell line was obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA). PC12 cells are a neuronal cell line derived from a rat pheochromocytoma, and these cells are often used as a model for neurons and a model for hypoxia because of their sensitivity to hypoxia. As described previously (19), briefly, cells were cultured in 75-cm² tissue culture flasks (Corning, NY, USA) in Dulbecco's modified Eagle medium (DMEM) (Gibco [Thermo Fisher Scientific]) and were maintained in DMEM containing 5% newborn calf serum and 10% horse serum supplemented with 100 U/mL penicillin and 100 pg/mL streptomycin in a moist atmosphere of 5% CO₂ at 37°C. Experiments were performed 1–2 d after plating.

Luciferase Expression in ODD-Luc Mice

ODD-Luc mice were given an intraperitoneal injection of 5-HMF, CoCl₂, ferulic acid, coumarin, quercetin or vehicle. One hour later, luciferin (50 mg/kg), which is a substrate of luciferase, was intraperitoneally injected into each mouse. Then, 5 min later, the mice were anesthetized by isoflurane for 5 min and then placed in a light-tight chamber equipped with an IVIS imaging camera (IVIS 50; Xenogen, Alameda, CA, USA). Photons were collected for a period of 10–30 s, and images were obtained and analyzed by using the Living Image software (Xenogen).

Hypoxia Treatment and Animal Survival

C57BL/6J mice were used to detect the effect of 5-HMF or VC on animal survival under a hypoxia environment. The mice were pretreated with 5-HMF (100 mg/kg), VC (140 mg/kg) or vehicle (normal saline) by intraperitoneal injection for 1 h and then supplemented with vehicles or VC (140 mg/kg) for another 1 h. After that, they were placed in a hypoxia chamber (DYC-DWI; Guizhou Fenglei, Guizhou, China) containing 20% O₂. For sublethal acute hypoxia, the oxygen levels were decreased at a velocity of approximately 1–2% O₂/min and reached 6% O₂ within 10 min. After exposed to hypoxia, animal survival was recorded. The survival time was defined with 10 min as the threshold value. The mice that lived over 10 min under hypoxia were considered survivors. Survival rate (%) = the number of mice that lived over 10 min/total number of mice.

Western Blot Analysis

The cortices and kidneys of ODD-Luc mice or PC12 cells were harvested, and the total proteins were extracted with radioimmunoprecipitation assay lysis buffer, which contained 50 mmol/L Tris-HCl (pH 7.4), 1% NP-40, 0.25% Nadeoxycholate, 150 mmol/L NaCl and 0.1% sodium dodecyl sulfate (SDS) supplemented with protease inhibitors (Roche, Indianapolis, IN, USA) and 1 mmol/L PMSF. The lysates were quantified by using a protein assay kit (Bio-Rad Laboratories, Hercules, CA, USA), separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to a polyvinylpyrrolidone difluoride membrane (Millipore, Billerica, MA, USA). After blocking with 5% nonfat dry milk in Tris-buffered saline with Tween-20 (TBST) at room temperature for 1 h, membranes were incubated with a HIF-1 α mouse monoclonal antibody (1:1,000; Novus Biologicals) and with a β -actin mouse monoclonal antibody (1:5,000; Sigma-Aldrich) in TBST, which contained 5% nonfat dry milk, at 4°C overnight. This overnight incubation was followed

by incubation with a goat anti-mouse secondary antibody, which was conjugated with horseradish peroxidase (1:1,000; Santa Cruz Biotechnology, Santa Cruz, CA, USA) in TBST, which contained 5% nonfat dry milk. Immune complexes on the membrane were visualized by using an enhanced chemiluminescence (ECL) detection system (Amersham Biosciences, Piscataway, NJ, USA). β -actin was used as a loading control. Experiments were repeated at least three times. Scans were used for semiquantitative analysis.

Immunofluorescence Staining

Cells that were grown on glass coverslips were fixed with methanol at –20°C for 20 min and permeabilized with 0.5% Triton X-100 at room temperature for 30 min. After blocking nonspecific binding with 5% normal goat serum in PBS-0.3% Triton X-100 at room temperature for 1 h, the cells were incubated with an anti-HIF-1 α antibody (1:500; Novus Biologicals). The primary antibody was detected by incubation with an Alexa Flour 594 goat anti-mouse secondary antibody (1:500; Molecular Probes [Thermo Fisher Scientific]) at room temperature for 1 h. All incubations were performed in a humidified chamber. The cells were visualized and photographed by using a confocal microscope (Leica TCS SP5; Leica, Heidelberg, Germany).

Reverse-Transcription-Polymerase Chain Reaction (RT-PCR)

For the analysis of *VEGF* mRNA, total RNA isolated from the cortices and kidneys of ODD-Luc mice was prepared by using the TRIzol[®] reagent (Invitrogen [Thermo Fisher Scientific]) according to the manufacturer's instructions. Total RNA (2 μ g) was amplified by a two-step protocol by using M-MLV reverse transcriptase and Taq polymerase. The PCR products were amplified by using the following program: primers for *VEGF* (forward: 5'-GACCCTGGTGGACATCTT-3', reverse: 5'-TAGTTCGCGAAACCCTGA-3') were used to yield a 452-base pair (bp) product for 28 cycles at 58°C; 18S rRNA primers (forward: 5'-GCCCG

AAGCGTTTACTTTGAA-3', reverse: 5'-GGTGAGGTTTCCCGTGTGA-3') were used to yield a 459-bp product for 28 cycles at 56°C. The mRNA expression level was semiquantified relative to the endogenous expression level of 18S rRNA as an internal control. PCR products were separated on a 1.5% (w/v) agarose gel and visualized by ethidium bromide staining. The images were acquired by using a MultiImage Light Cabinet (Alpha Innotech, San Leandro, CA, USA).

Quantitative Real-time PCR (qRT-PCR)

The mRNA levels of *HIF-1 α* and *β -actin* were measured using Power SYBR[®] Green (Applied Biosystems [Thermo Fisher Scientific]) and an ABI StepOnePlus Real-Time PCR System (Applied Biosystems [Thermo Fisher Scientific]) according to the manufacturer's instructions. Total RNA was isolated from PC12 cells by using the TRIzol[®] Reagent and performed according to the manufacturer's protocol. An aliquot of 1 μ g total mRNA was reverse-transcribed at 42°C for 1 h in a 10- μ L reaction mixture. The qRT-PCR was performed in triplicate by using Power SYBR[®] Green and a real-time PCR system. The following *HIF-1 α* primers were used: forward 5'-CCGCAGTGTGGCTAC AAGAA-3', reverse 5'-GATGAGGAAT GGGTTCACAAATC-3', 67 bp. The results were normalized by using *β -actin* as the internal control, with the following primers: forward 5'-CCAGTTCGCC ATGGATGAC-3', reverse 5'-ATGCC GGAGCCGTTGTC-3', 87 bp. Calculations were based on the "delta-delta method" by using the following equation: R (ratio) = $2^{-[\Delta\text{CT}]_{\text{sample}} - \Delta\text{CT}_{\text{control}}}$ (21). The data were expressed as fold changes of the treatment groups relative to the control. β -actin was amplified as an internal control for qRT-PCR analysis.

Ultraviolet-Visible Spectroscopy Analysis

To detect the interaction of 5-HMF with Fe²⁺ (FeSO₄), 2-OG or VC, an ultraviolet (UV)-visible spectrophotometer (Multiskan Go; Thermo Scientific [Thermo Fisher Scientific]) was used for the analy-

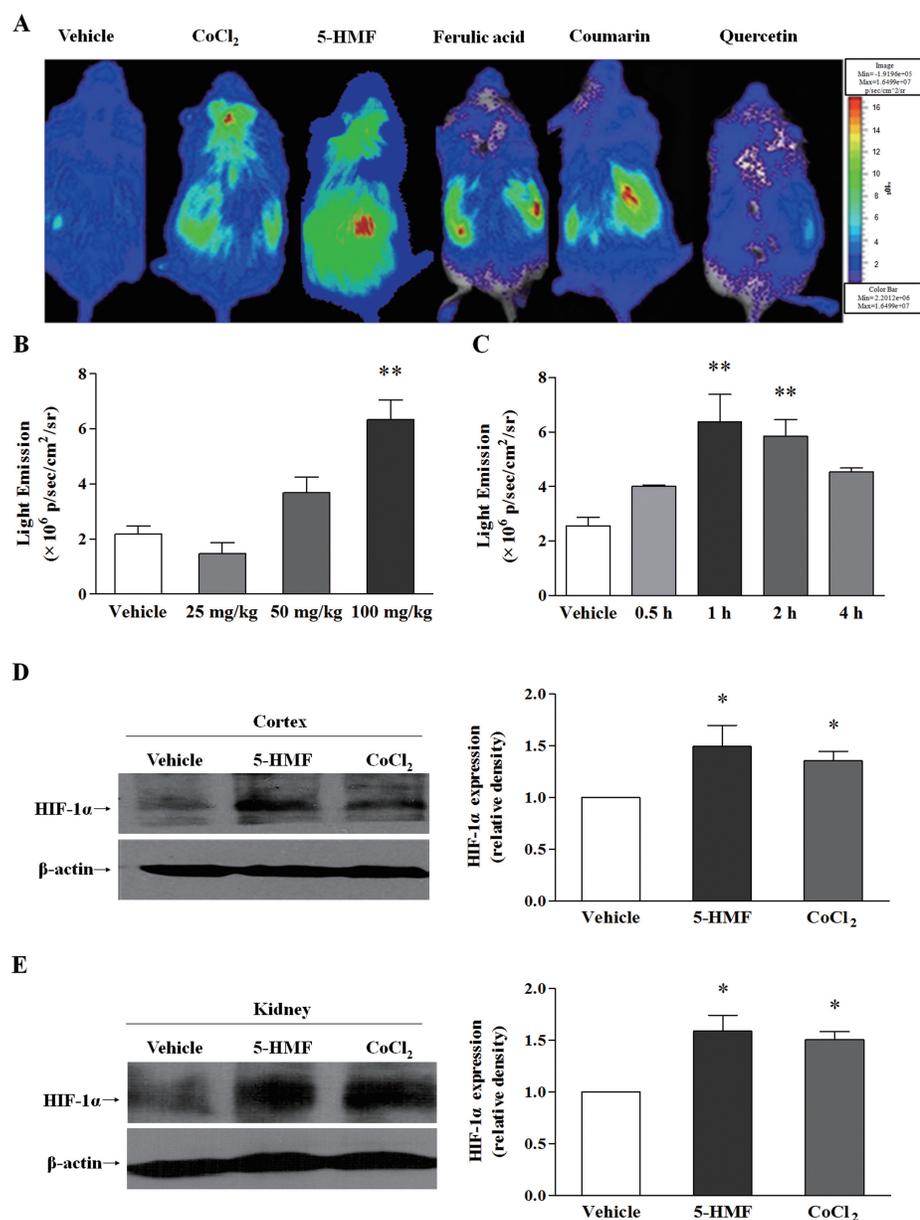


Figure 1. The stabilization effect of 5-HMF on HIF-1 α in ODD-Luc mice. (A) Comparison of the bioluminescence among anti-oxidative agents. The intensity of bioluminescence indicates the degree of HIF-1 α stability. The ODD-Luc mice were given an intraperitoneal injection of vehicle (normal saline), CoCl₂ (125 mg/kg), 5-HMF (100 mg/kg), ferulic acid (100 mg/kg), coumarin (100 mg/kg) or quercetin (100 mg/kg). The color bar indicates photons/(s/cm²/steradian), with minimum and maximum threshold values. (B, C) Determination of the dose and time effect of 5-HMF on HIF-1 α stability. The left panel shows the bioluminescence intensity 1 h after the mice were treated with different concentrations of 5-HMF. The right panel shows the bioluminescence intensity at 0.5, 1, 2, and 4 h after the mice were treated with 100 mg/kg of 5-HMF. The data that are shown are the means \pm SE of three independent experiments. ** $p < 0.01$ compared with the vehicle. (D, E) Western blot analysis of HIF-1 α protein expression in the cortex and kidney tissues of ODD-Luc mice. Whole-cell lysates were subjected to an immunoblot assay for HIF-1 α and β -actin. β -actin was used as an internal control. The values were normalized to the vehicle levels. The results are the means \pm SE of four independent experiments. * $p < 0.05$ compared with the vehicle.

ses. The solution of 20 μ mol/L 5-HMF and 0, 2.5, 5, 10, 15 or 20 μ mol/L FeSO₄, 2-OG or VC was mixed and incubated for 2 min at room temperature. Spectra were collected in the range of 200- to 500-nm wavelengths. The maximum UV absorptivity of 5-HMF is 285 nm.

To determine the VC content in the cytoplasm, sample preparation and measurements, as described previously in the literature (22,23), were as follows: briefly, after PC12 cells were treated with 5-HMF, the cells were permeabilized with 0.5% Triton X-100 in phosphate-buffered saline (PBS) for 10 min, and the supernatant was collected. Then, 200 μ L of the supernatant was added to 1 mL 0.1 mmol/L NaH₂PO₄ (pH 5.6) and 750 μ L ultrapure water and mixed. The absorbance at 265 nm (A_{265}) was measured by using a spectrophotometer, followed by incubation with ascorbate oxidase (0.4 U, pH 5.6) for 3 min and measured again. The decrease in A_{265} was monitored. The VC content was calculated based on the decrease in A_{265} .

High-Performance Liquid Chromatography (HPLC) Analysis of 5-HMF Content

The stock solution of 5-HMF (4 μ mol/L) was prepared by dissolving 25.22 mg 5-HMF in 50 mL ultrapure water and stored at 4°C. Five concentrations of 5-HMF standard solutions (0.02, 0.10, 0.50, 2.52 and 12.61 mg/100 mL) were obtained by appropriately diluting the stock solution with mobile phase and were used to evaluate the linearity of the HPLC data (HP1100; Agilent, Burnsville, MN, USA). Chromatographic separation was performed by using a Diamonsil C18 column (250 \times 4.6 mm inside diameter [i.d.], 5 μ m; Dikma, Lake Forest, CA, USA). The mobile phase was a mixture of ultrapure water to acetonitrile (95:5, v/v) and was delivered at a flow rate of 1.0 mL/min. The detection wavelength was 285 nm.

Statistical Analysis

Statistical analysis was performed by using the SPSS software. All the data were expressed as the means \pm standard

error (SE). A comparison between two samples was performed using a two-tail Student *t* test. *p* < 0.05 was considered statistically significant.

RESULTS

5-HMF Increased HIF-1 α Stability in a Dose- and Time-Dependent Manner in ODD-Luc Transgenic Mice

Using ODD-Luc transgenic mice, which are a useful model to probe the stabilization of HIF-1 α , we compared the effect of 5-HMF and the other three confirmed compounds that can stabilize HIF-1 α . Cobalt chloride (CoCl₂), which is an iron antagonist, was used as a positive control. Compared with ferulic acid, coumarin and quercetin, 5-HMF caused stronger bioluminescence in the brain and kidney areas (Figure 1A), which indicated a stronger role of 5-HMF in stabilizing HIF-1 α . In addition, 5-HMF stabilized HIF-1 α in a dose- and time-dependent manner in ODD-Luc mice (Figures 1B, C). To further verify this effect of 5-HMF on HIF-1 α stability, we measured HIF-1 α protein expression in the cortices (Figure 1D) and kidneys (Figure 1E) of ODD-Luc mice by Western blot. Consistent with the result of the bioluminescent assay, 5-HMF accumulated the HIF-1 α protein in both tissues.

5-HMF Promoted HIF-1 α Stability and Nuclear Translocation in PC12 Cells

In hypoxia-sensitive PC12 cells, the role of 5-HMF in accumulating HIF-1 α was further determined. Through comparing HIF-1 α protein levels 1 h after cells were treated with different concentrations of 5-HMF, we identified the role of 100 μ g/mL 5-HMF in the accumulation of HIF-1 α (Figure 2A). To identify whether this accumulating effect of 5-HMF on the HIF-1 α protein is due to promoting HIF-1 α stability or to increasing HIF-1 α transcription, we measured the mRNA expression of *HIF-1 α* by real-time PCR. Although there was a subtle trend of increasing *HIF-1 α* mRNA with increasing 5-HMF dosage, there were no significant differences

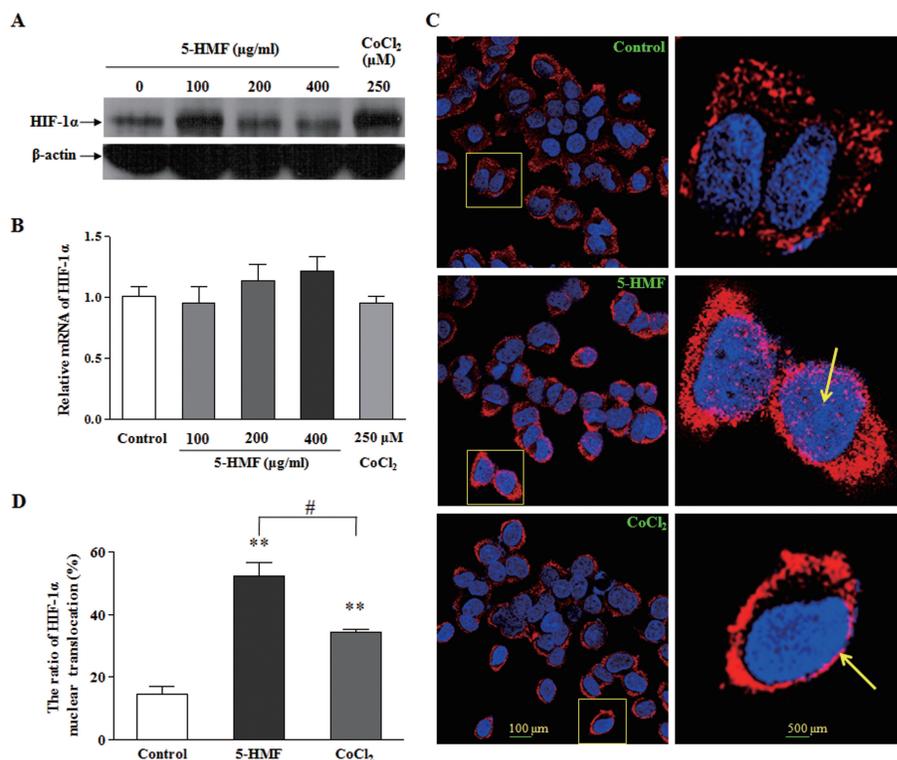


Figure 2. The increased stability and nuclear translocation of HIF-1 α by 5-HMF in PC12 cells. (A) Effects of 5-HMF on the levels of the HIF-1 α protein. PC12 cells were exposed to the indicated concentrations of 5-HMF or CoCl₂ for 1 h. The representative immunoblots are shown. (B) Effects of 5-HMF on the levels of *HIF-1 α* mRNA. The *HIF-1 α* mRNA levels were determined by qRT-PCR analysis after cells were treated for 1 h. The relative expressions were normalized to the control level. (C) Effects of 5-HMF on the nuclear translocation of HIF-1 α . The distribution of the HIF-1 α protein was analyzed with immunofluorescence staining after the cells were treated with 100 μ g/mL 5-HMF for 1 h. The cells were labeled for HIF-1 α (red) and counterstained with DAPI (blue), which was used to label nuclei. The arrowheads indicate the nuclear localization of HIF-1 α , which was stained pink. The image at the right panel is the enlarged image in the frame at the left. (D) Statistical analysis of the ratio of HIF-1 α nuclear translocation. The ratio was calculated in terms of the following formula: HIF-1 α nuclear translocation (%) = (the number of cells with pink/the total number of cells with red) \times 100. The results are expressed as the means \pm SE from three independent experiments. ***p* < 0.01 compared with the control; #*p* < 0.05 for comparisons between the indicated groups.

compared with the control. Therefore, 5-HMF did not cause an obvious increase in the *HIF-1 α* mRNA level (Figure 2B). This result suggests that the higher level of the HIF-1 α protein that was induced by 5-HMF was due to the stabilization of the HIF-1 α protein in normoxia rather than by the promotion of its *de novo* protein synthesis.

By immunofluorescence staining, we observed that HIF-1 α gathered to the nucleus and partly entered into the nucleus in the cells that were treated with

5-HMF; in contrast, HIF-1 α was scattered in the cytoplasm in the control (Figure 2C). By counting the number of cells where HIF-1 α entered into their nuclei and the total number of cells that expressed HIF-1 α , we obtained the ratio of HIF-1 α nuclear translocation. The statistical data revealed that 5-HMF significantly increased the ratio of HIF-1 α nuclear translocation (Figure 2D). In short, the above results show that 5-HMF promotes HIF-1 α stability and its nuclear translocation.

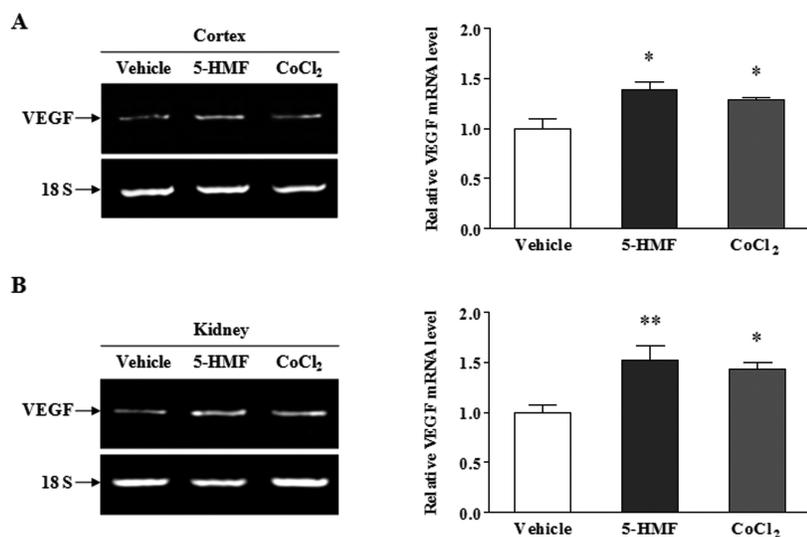


Figure 3. The enhanced *VEGF* mRNA expression by 5-HMF in ODD-Luc mice. (A, B) Effects of 5-HMF on the *VEGF* mRNA expression in the cortex and in the kidney. The transcriptional activity of HIF-1 was estimated by *VEGF* mRNA expression. The *VEGF* mRNA levels were analyzed by semiquantitative RT-PCR. Tissues were harvested for RT-PCR 1 h after the ODD-Luc mice were given an intraperitoneal injection of vehicle (normal saline), 5-HMF (100 mg/kg) or CoCl₂ (125 mg/kg). 18S rRNA was used as an internal control. The values were normalized to the vehicle levels. The data are representative of four different experiments and are expressed as the means ± SE. * $p < 0.05$, ** $p < 0.01$, compared with the vehicle.

5-HMF Increased HIF-1 Transcriptional Activity by the Induction of *VEGF* mRNA Expression in ODD-Luc Transgenic Mice

To determine the effect of 5-HMF on HIF-1 transcriptional activity after stabilizing HIF-1 α , we measured the mRNA level of *VEGF*, which is an HIF-1 target gene, to assess HIF-1 transcriptional activity. After the total mRNA was extracted from the cerebral cortex and kidney of ODD-Luc mice that were treated with 100 mg/kg 5-HMF for 1 h, the expression of *VEGF* mRNA was measured by RT-PCR. Compared with the vehicle, the levels of *VEGF* mRNA in the cortex (Figure 3A) and kidney (Figure 3B) were elevated by 5-HMF, which was consistent with the case of the HIF-1 α protein. This result suggests that 5-HMF increased HIF-1 transcriptional activity.

5-HMF Reacted with VC *In Vitro* and Reduced the VC Content in PC12 Cells

How did 5-HMF increase the stability of HIF-1 α ? HIF-1 α stability depends on

the activity of PHDs, which is determined by the content of the substrates (O₂, 2-OG) or the cofactors (Fe²⁺, VC) of PHDs. Because 5-HMF increased HIF-1 α stability in normoxia, we considered whether 5-HMF inhibited the activity of PHDs through interacting with 2-OG, Fe²⁺ or VC. Using UV spectrophotometry, we discovered that different concentrations of Fe²⁺ or 2-OG did not affect the maximum absorbance wavelength (λ_{\max}) of 5-HMF; however, with increasing VC concentrations, the λ_{\max} of 5-HMF decreased (Figure 4A). The correlation curve of the concentration–absorbance difference (Δ Abs) revealed that there was no correlation between 5-HMF and Fe²⁺ or 2-OG; however, there was a significant correlation between 5-HMF and VC (Figure 4B), which implied a biochemical reaction between 5-HMF and VC. Thus, we wondered whether 5-HMF could enter cells and affect their VC content. After PC12 cells were treated with 5-HMF for 1 h, the intracellular content of 5-HMF was detected using HPLC. The results showed that the intracellular contents of

5-HMF increased with the extracellular concentrations of 5-HMF (Figure 4C). This result suggests that 5-HMF can enter cells in a dose-dependent fashion. Subsequently, we further detected the VC content in PC12 cells by using UV spectrophotometry after the cells were treated with 5-HMF. The VC content in PC12 cells was significantly reduced by 5-HMF (Figure 4D). These results suggest that 5-HMF reacted with VC in the cells and, therefore, reduced the VC content.

Increased HIF-1 α Stability by 5-HMF Was Reversed by VC Supplementation in PC12 Cells and in ODD-Luc Mice

To verify that the increased HIF-1 α stability caused by 5-HMF was due to a reduction in the VC content, we measured the hydroxylation level of HIF-1 α Pro-564, which indirectly reflects the activity of PHDs, and the protein level of HIF-1 α in PC12 cells by Western blot. 5-HMF decreased the level of HIF-1 α Pro-564 hydroxylation, whereas VC supplementation returned the protein level of HIF-1 α to the control level (Figure 5A). Similarly, the accumulation of HIF-1 α induced by 5-HMF was also attenuated by VC supplementation (Figure 5B). By using ODD-Luc mice, we observed that 5-HMF induced an increase in bioluminescence, and VC supplementation resulted in a decrease in bioluminescence compared with the effect of 5-HMF (Figure 5C). Altogether, these results indicate that the role of 5-HMF in stabilizing HIF-1 α can be reversed by VC supplementation, which suggests that the stabilization of HIF-1 α by 5-HMF is mediated by a reduction in the VC content.

Increased Survival of Mice by 5-HMF Under Hypoxia Was Reversed by VC Supplementation

To elucidate the effect of increased HIF-1 α stability by 5-HMF on mice survival under hypoxia, we compared the survival of mice that were treated with 5-HMF or that were supplemented with VC. The survival time of mice under hypoxia was extended by 5-HMF; in contrast, VC supplementation obviously decreased the survival time (Figure 6A). Simultaneously, the

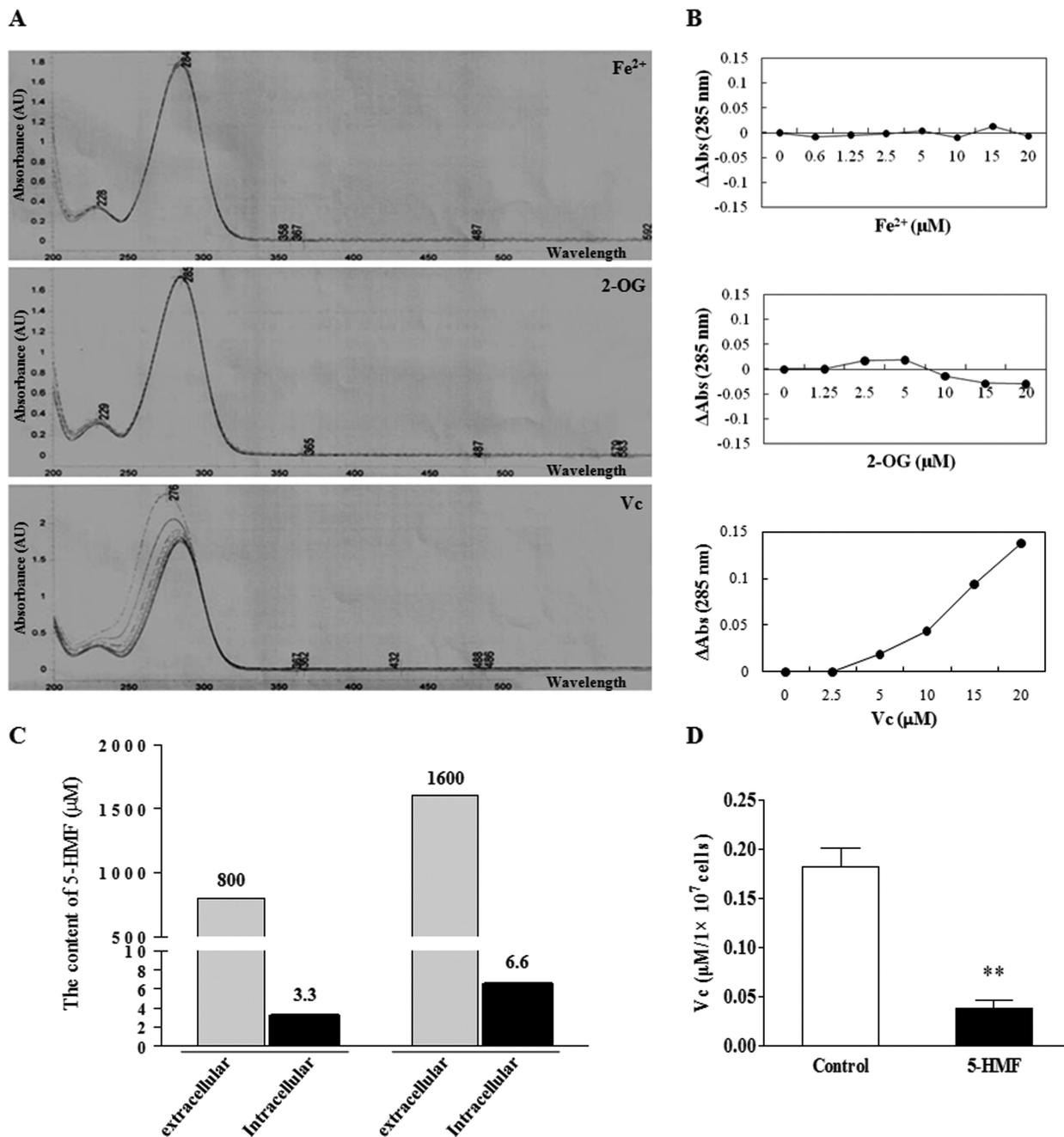


Figure 4. The reaction of 5-HMF and VC *in vitro* and, thereby, a reduction in the VC content in PC12 cells. (A) The UV-visible absorption spectra of 5-HMF. A total of 20 $\mu\text{mol/L}$ 5-HMF in 10 mmol/L phosphate buffer, pH 7.4, was titrated with increasing concentrations (0, 2.5, 5, 10, 15 and 20 $\mu\text{mol/L}$) of Fe^{2+} , 2-OG or VC *in vitro*. After incubation for 2 min, the absorption spectra of 5-HMF was obtained from 200 to 500 nm by UV-visible spectroscopy analysis. The absorption peak of 5-HMF is 285 nm. (B) The correlation curves of concentration and absorbance difference compared with 285 nm (ΔAbs 285 nm). There is no correlation between 5-HMF and Fe^{2+} or 5-HMF and 2-OG; however, there is a positive correlation between 5-HMF and VC, which indicates the reaction of 5-HMF and VC. (C) Determination of the content of 5-HMF in the intracellular cytoplasm. The content of 5-HMF in PC12 cells was detected by HPLC after the cells were exposed to 5-HMF in the medium for 1 h. 5-HMF was detectable in the cytoplasm, and the content increased with increasing concentrations of extracellular 5-HMF. (D) The reduction of VC content by 5-HMF. The VC content was detected by UV-visible spectroscopy analysis after the cells were exposed to 5-HMF (100 $\mu\text{g}/\text{mL}$) for 1 h. The data are representative of four different experiments and are expressed as the means \pm SE. ** $p < 0.01$, compared with the control.

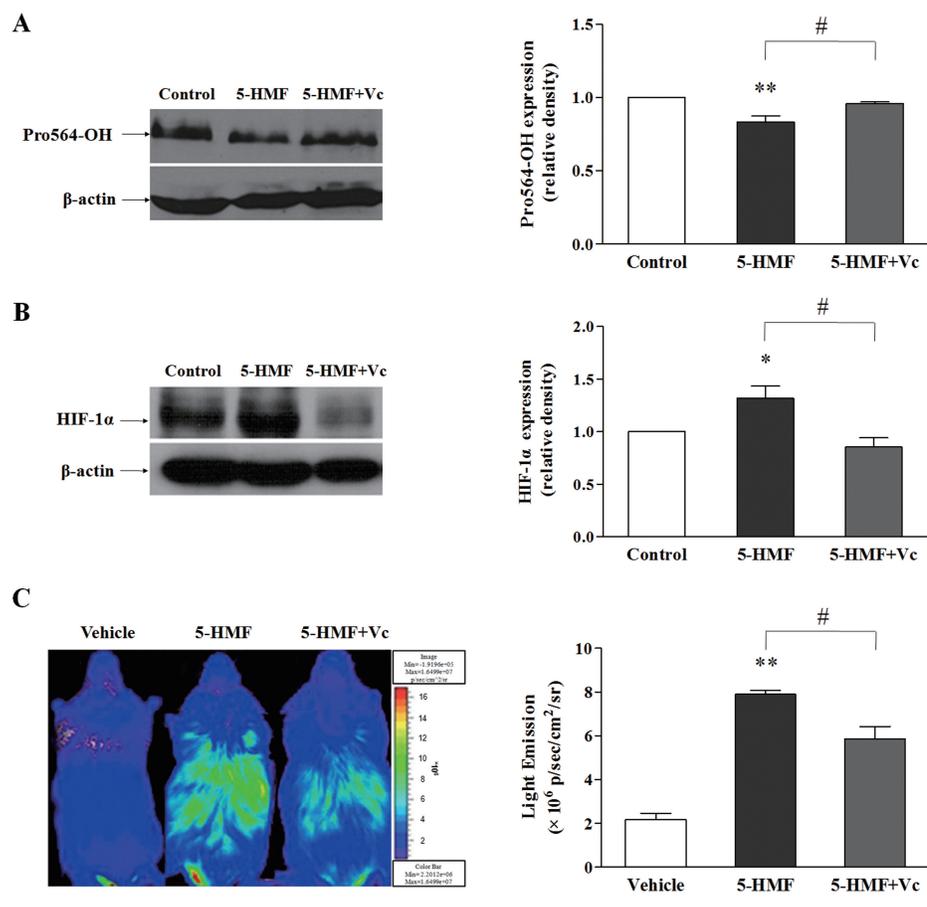


Figure 5. VC reversed the stabilization of HIF-1 α , which was caused by 5-HMF, in PC12 cells and in ODD-Luc mice. (A, B) Western blot analysis of the hydroxylation of HIF-1 α Pro-564 and the HIF-1 α protein expression. The levels of HIF-1 α hydroxylation and HIF-1 α protein expression were measured by Western blot after PC12 cells were exposed to 100 μ g/mL 5-HMF or culture medium for 1 h, followed by supplementation with culture medium or with 140 μ g/mL VC for another 1 h. The values were normalized to the control levels. The data that are shown are the means \pm SE of four independent experiments. * p < 0.05, ** p < 0.01, compared with the control; # p < 0.05 for comparisons between the indicated groups. (C) Bioluminescence analysis of HIF-1 α stability in ODD-Luc mice. After the ODD-Luc mice were given an intraperitoneal injection of 5-HMF (100 mg/kg) or vehicle (normal saline) for 1 h, the mice were supplemented with VC (140 mg/kg) or with normal saline for another 1 h. The data shown are the means \pm SE of four independent experiments. ** p < 0.01 compared with the vehicle, # p < 0.05 for comparisons between the indicated groups. The color bar indicates photons/(s/cm²/steradian), with minimum and maximum threshold values.

survival rate of mice under hypoxia was significantly increased by 5-HMF; however, VC supplementation apparently reduced the survival rate (Figure 6B). Thus, it is clear that the increased survival of mice induced by 5-HMF under hypoxia is correlated with the increased stability of HIF-1 α by a reduction in the VC content.

Taken together, we demonstrated the interaction of 5-HMF with VC and the

resulting reduction in VC content, which inhibited the activity of PHDs and ultimately stabilized HIF-1 α through blocking HIF-1 α hydroxylation-mediated degradation (Figure 7).

DISCUSSION

We first reported the role of 5-HMF against hypoxia in 2011 (5,6); however, the potential mechanism had not yet

been elucidated. By using ODD-Luc mice, which are an ideal model for identifying and characterizing small molecular inhibitors of PHD (19,25), we found the role of 5-HMF in stabilizing HIF-1 α . The ODD-Luc mice were generated by Safran *et al.* in 2006 (24). The mice provide a rapid noninvasive measurement of possible agonists of HIF-1 activity and are considered an ideal model that is suited for identifying and characterizing small molecular inhibitors of PHDs that would increase HIF-1 activity (25). We tested several compounds to find out the best one stabilizing HIF-1 α using these ODD-Luc mice. Ferulic acid and coumarin increased the activity of HIF-1 α in kidney but not in brain, and quercetin hardly played a role in stability of HIF-1 α . Compared with the other three compounds, 5-HMF displayed a more significant role in stabilizing HIF-1 α . The stabilized HIF-1 α was concentrated in the kidney area and part of the brain regions, and the stabilization was dose and time dependent. These results indicate that 5-HMF shows not only a strong role in stabilizing HIF-1 α , but also a relatively wide distribution. In particular, the increased HIF-1 α activity in brain hints that 5-HMF can cross the blood–brain barrier, which suggests a more promising clinical application. To verify the effect of 5-HMF on HIF-1 α , we further measured the protein expression of HIF-1 α in the cortex and kidney of ODD-Luc mice by Western blot assay. In both the cortex and the kidney, 5-HMF increased HIF-1 α expression. Thus, we obtained a preliminary conclusion that 5-HMF improved HIF-1 α stability at the animal level.

By using hypoxia-sensitive PC12 cells, we confirmed the role of 5-HMF in stabilizing HIF-1 α and explored the potential HIF-1 α stabilization pathway of 5-HMF. The HIF-1 α protein was accumulated by 5-HMF in a dose-dependent manner. To determine whether this effect was due to the enhanced HIF-1 α stability or due to the increased protein *de novo* synthesis, we detected the level of HIF-1 α mRNA by quantitative real-

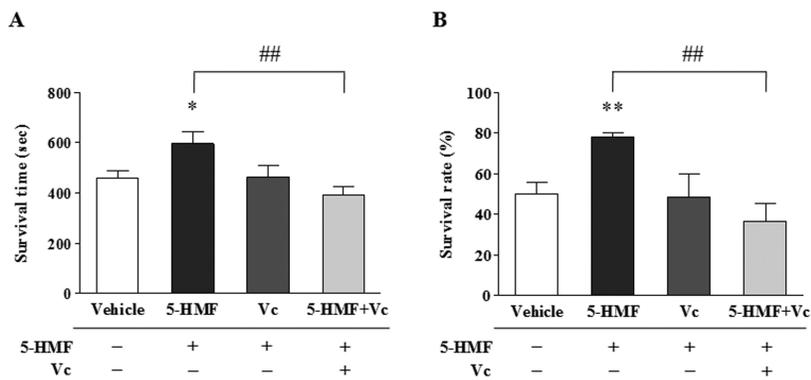


Figure 6. VC reversed the increased mice survival that was caused by 5-HMF under hypoxia. (A, B) The survival time and the survival rate under hypoxia. After the C57BL/6J mice were given an intraperitoneal injection of 5-HMF (100 mg/kg), VC (140 mg/kg) or vehicle (normal saline) for 1 h, which was followed by supplementation with vehicles or with VC (140 mg/kg) for another 1 h, the mice were exposed to sublethal acute hypoxia (6% O₂) in a hypoxia chamber, and their survival time was recorded. The data that are shown are the means \pm SE of three independent experiments. * p < 0.05, ** p < 0.01, compared with the vehicle; ## p < 0.01 for comparisons between the indicated groups.

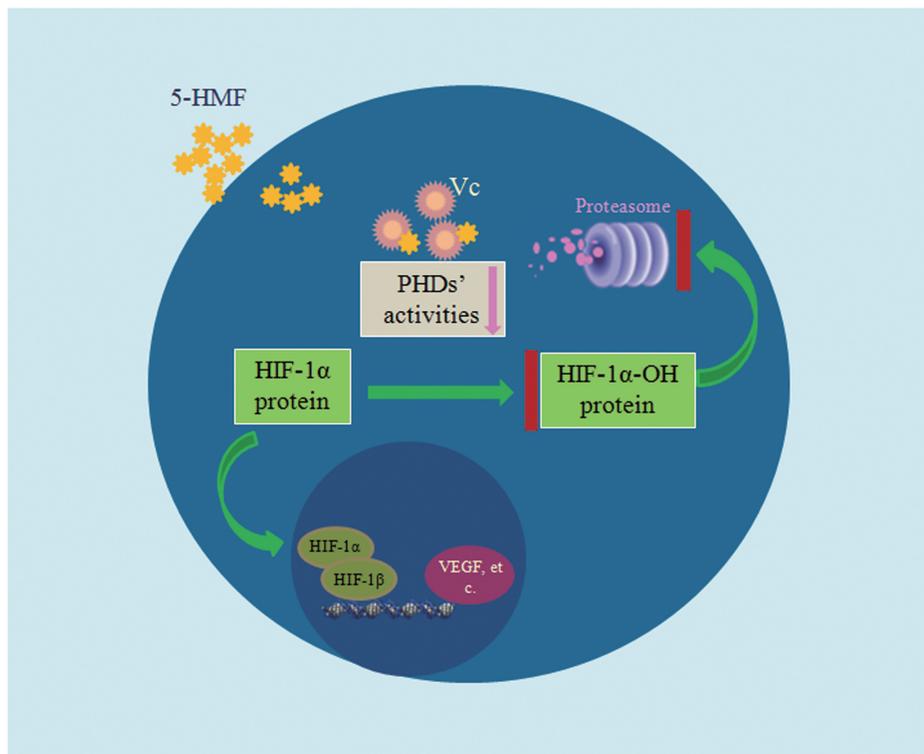


Figure 7. Model depicting the role of 5-HMF in stabilizing HIF-1 α . 5-HMF enters cells in a dose-dependent manner. Then, 5-HMF rapidly interacts with VC and thereby reduces the VC content. PHD activities are declined because of the reduction in VC. HIF-1 α hydroxylation is inhibited because of the reduction of PHD activity, and thus, HIF-1 α cannot be degraded through the ubiquitin-proteasome pathway. Therefore, the HIF-1 α protein is stabilized and then translocates into the nucleus. In the nucleus, HIF-1 α combines with HIF-1 β to form the heterodimeric transcription factor HIF-1, which regulates the transcription of its downstream genes, such as *VEGF*.

time PCR after the cells were treated with 5-HMF. Although there were no significant differences between 5-HMF and the control, there was a subtle trend of increasing *HIF-1 α* mRNA with increasing 5-HMF dosage. This result could indicate that the action of 5-HMF may not solely affect PHD function and may increase HIF-1 α levels.

In addition to regulating HIF-1 α stability, 5-HMF was also involved in promoting the nuclear translocation of HIF-1 α . One viewpoint is that the rapid nuclear translocation of HIF-1 α is an efficient way to escape from proteasomal degradation (26); another viewpoint is that nuclear translocation is not necessary for HIF-1 α stabilization because both “nuclear” and “cytoplasmic” proteasomes are fully competent for HIF-1 α degradation in an oxygen-dependent manner (27). We believe that the nuclear translocation of HIF-1 α is one active mechanism of HIF-1 α and is a necessary process to form a heterodimer with HIF-1 β in the nucleus. Perhaps HIF-1 α cannot be degraded in the nucleus once the heterodimer is formed. Regardless, the nuclear translocation of HIF-1 α is a premise to form the heterodimeric transcription factor HIF-1. In view of this result, we tested HIF-1 α translocation by using immunofluorescence staining. By confocal scanning, we directly observed the nuclear translocation of HIF-1 α , which was promoted by 5-HMF. The increased ratio of HIF-1 α nuclear translocation suggested the enhanced activity of HIF-1 α . As some literature previously reported, once HIF-1 α is stable, this protein can rapidly translocate into the nucleus and activate the transcriptional activity of HIF-1 (28,29).

To further assess whether the transcription activity of HIF-1 is activated by 5-HMF, we measured the mRNA expression of *VEGF*, which is a downstream gene of HIF-1, in the cortex and the kidney of ODD-Luc mice by RT-PCR. Consistent with the above results, 5-HMF improved the level of *VEGF* mRNA in both the cortex and the kidney. Thus far, we can say that 5-HMF promotes HIF-1 α

stability, which therefore leads to the enhanced transcriptional activity of HIF-1.

The stability of HIF-1 α is determined by the activity of PHDs. To determine the mechanism by which 5-HMF stabilizes HIF-1 α , we analyzed the interaction of 5-HMF with Fe²⁺, 2-OG or VC *in vitro*, which determines the activity of PHDs. The result showed that 5-HMF did not react with Fe²⁺ or 2-OG but did react with VC. However, we still do not know the product that is generated from the reaction. In addition, we demonstrated that 5-HMF could enter the cells in a dose-dependent manner and reduce the intracellular VC content. This result suggests that 5-HMF reacted with VC in cells and thereby reduced the VC content.

Because 5-HMF reduced the VC content, could 5-HMF affect PHD activity? As we know, increased PHD activity can cause the prolyl hydroxylation of HIF-1 α and its subsequent degradation. In view of the difficulty in measuring PHD activity directly, we tested the level of hydroxylated 564 prolyl residues of HIF-1 α to indirectly reflect the activity of PHDs. We detected the reduced prolyl hydroxylation of HIF-1 α in those cells that were treated with 5-HMF, whereas VC supplementation reversed this effect of 5-HMF. Next, we verified the reverse effect of VC on 5-HMF-induced HIF-1 α stability separately in PC12 cells and in the ODD-Luc mice. Thus, we can speculate that PHD activity was inhibited by 5-HMF, which might be achieved through a reduction in VC content by the interaction of 5-HMF with VC, and thus, the stability of HIF-1 α was improved. A recent finding that VC plays a significant role in supporting HIF-hydroxylase function and thereby modulates the cell survival response was consistent with our speculation (30).

To verify the effect of HIF-1 α stability on the protection against hypoxia, we used C57BL/6J mice to test the survival of mice under hypoxia. The survival time and the survival rate of the mice that were pretreated with 5-HMF both increased, most likely because of the improved HIF-1 α stability by 5-HMF. VC

supplementation reversed this effect exerted by 5-HMF, which illustrated the role of HIF-1 α stability in hypoxia protection. This role is contrary to the role of VC in tumors, in which HIF-1 is an important regulator of the adaptation of the tumor to its hypoxic microenvironment, and VC administration can significantly reduce tumor growth rates (31). This observation indicates that the same mechanism can lead to opposite effects due to different pathophysiological states. In this study, pretreatment with 5-HMF increased the stability of HIF-1 α that subsequently translocated to the nucleus and activated the transcription of its downstream genes, including *VEGF*, *EPO*, *GLUTs*, and so on, which can enhance hypoxia tolerance (16–18,32) and ultimately increased survival time and survival rate of the mice. The reverse effect of VC supplementation on survival further confirmed that the role of 5-HMF in promoting survival under hypoxia was probably mediated by regulation on the HIF-1 α pathway.

CONCLUSION

In summary, we identified a molecular mechanism by which 5-HMF increases HIF-1 α stability in normoxia, which may play a critical role in the protection of 5-HMF against hypoxia. The regulation of HIF-1 α stability by 5-HMF is an important pathophysiological mechanism that may provide a novel therapeutic approach to hypoxic diseases.

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DISCLOSURE

The authors declare that they have no competing interests as defined by *Molecular Medicine*, or other interests that might be perceived to influence the results and discussion reported in this paper.

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